BRIEF REPORT

Detection of *Mycoplasma canadense* and *Mycoplasma californicum* in dairy cattle from Argentina

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**Abstract**

Different species of *Mycoplasma* can affect bovine cattle, causing several diseases. PCR sequencing and further analysis of the 16S-23S rRNA ITS region have shown a significant interspecies variability among *Mollicutes*. Sixteen suspected isolates of *Mycoplasma* spp. obtained from milk samples from dairy herds were amplified (16S-23S rRNA ITS region). Fourteen out of those 16 suspected *Mycoplasma* spp. isolates were PCR-positive. To confirm the identity of *Mycoplasma bovis*, these 14 isolates were tested by another species-specific PCR. Seven of the isolates rendered a positive result. The products of 16S-23S rRNA ITS PCR from one isolate that was identified as *M. bovis* and from two other isolates, identified as non-*M. bovis* were randomly selected, sequenced and analyzed. The three sequences (A, B and C) showed 100% similarity with *M. bovis*, *Mycoplasma canadense* and *Mycoplasma californicum* respectively.

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Different species of *Mycoplasma* can affect bovine cattle, causing several diseases. Mycoplasmas can cause clinical, subclinical or chronic intramammary infection affecting cattle of all ages and at any stage of lactation. In Argentina, *Mycoplasma bovis* was firstly reported in the year 2000 in a bovine mastitis outbreak in Buenos Aires province. Since then, *Mycoplasma* spp. has been frequently isolated but there are no literature reports about the identification of other *Mycoplasma* species affecting bovine cattle.

As in bacteria, in mycoplasmas the rRNA genes (16S-23S-5S) are separated by internal transcriber spacer (ITS) regions. Sequencing and analysis of the 16S-23S rRNA ITS region have shown a significant interspecies variability among *Mollicutes*. In fact, this region has been proposed as a complementary genetic marker for species identification of the genera *Mycoplasma*.4 In our laboratory, we routinely use the amplification of a fragment of the 16S-23S ITS region as a screening PCR test for *Mycoplasma* spp. identification from isolates or clinical samples from dairy cattle, in which *Mycoplasma* infection is suspected. Due to the novelty of our findings and the lack of information about the *Mycoplasma* species affecting dairy cattle in our country, the objective of this study is to report the detection of *Mycoplasma bovis*, *Mycoplasma canadense* and *Mycoplasma californicum* in dairy cattle from Argentina, by sequencing the 16S-23S rRNA ITS region of the isolates.

This work was performed at the Laboratory of Animal Pathology of the Faculty of Agronomy and Veterinary Sciences (UNRC, Rio Cuarto, Córdoba, Argentina), according to the international guidelines of the Council for International Organizations of Medical Sciences (CIOMS).

Culture and DNA extraction. Sixteen suspected isolates of *Mycoplasma* spp. obtained from milk samples from dairy herds were analyzed. Milk samples were taken from mammary quarters of cows with and without clinical mastitis and from bulk tank milk from herds from Córdoba and Buenos Aires provinces. These samples were cultured in Modified Hayflick’s medium plates at least 7 days at 36 +/- 1 ºC with 10% CO₂. *Mycoplasma* spp. suspected colonies were identified by daily examination of plates under a stereomicroscope. These colonies were picked up and inoculated into Modified Hayflick broth medium and incubated for 48 hs at 36 +/- 1 ºC with 10% CO₂. One ml of each culture was centrifuged at 8,000 × g for 10 min, pellets were washed twice with PBS solution and suspended in 150 μl of sterile purified water. DNA was extracted by boiling (10 min).

PCR and sequencing. DNA was subjected to PCR for 16S-23S rRNA ITS fragment amplification to identify *Mycoplasma* using the primers and PCR conditions previously described. Fourteen out of 16 suspected *Mycoplasma* spp. colonies were PCR-positive to 16S-23S rRNA ITS amplification reaction. The size of the amplicons varied approximately from 350 bp to 500 bp. To identify *M. bovis*, these 14 colonies were tested by another PCR, amplifying a fragment of the *uvrC* gene using the primers and conditions described by Subramanian et al. and Thomas et al. respectively. Seven of the colonies rendered a positive result.

PCR products from 16S-23S rRNA ITS amplification from one of the colonies identified as *M. bovis* (isolate A) and two other colonies identified as *Mycoplasma non-M. bovis* (isolates B and C) were randomly selected for further analysis. These three amplicons were purified (Puriprep-GP Kit, InBioHighway, Tandil, Argentina), quantified, and sequenced (ABI 3130xL; Applied Biosystems, Foster City, California) with the primers described by Harasawa et al. The sequences were visualized using the BioEdit software and were aligned against the database using nucleotide BLAST. In order to eliminate the flanking regions (corresponding to partial sequences of the 16S rRNA and 23SrRNA genes) the sequences were aligned using ClustalW. The 16S-23S rRNA ITS sequence obtained from isolate A showed 100% similarity with the same region of *Mycoplasma bovis* strains PG45 (CP002188.1), 70-213 (AY779747.1), Hubei-1 (CP002058.1),HB0801 (CP002058.1), HEK-FDA (JN644755.1), Madison (AY780798.1), and ATCC 25025 (AY765211.1). These results were in accordance with the identification of this isolate as belonging to *M. bovis* by PCR amplification of *uvrC*.

The 16S-23S rRNA ITS sequence obtained from isolate B showed 100% similarity with the same region of *Mycoplasma canadense* strains QMP-SRI-0053 (KC759701.1), QMP-SRI-0054 (KC771072.1), QMP-SRI-0052 (KC485347.1), 275C(DQ847417.1), ATCC 29418 (AY800341.1), and 466 (EU925158.1; DQ847418.1). The sequence alignment from strain C to *Mycoplasma californicum* strains S6 (DQ847428.1) and ATCC 33461 (AY736031.1) showed 100% similarity in both cases.

The present study confirms the presence of this pathogen in milk samples by sequencing the 16S-23S rRNA ITS region and also by the amplification of a species-specific gene *uvrC*; however, this case corresponded to an isolate from a cow with clinical mastitis from Córdoba province (data not shown).

It is worth noting that in Argentina, the presence of *M. canadense* and *M. californicum* has never been reported before. *M. canadense* and *M. californicum* together with *M. bovis*, *M. arginini*, *M. bovigenitalum*, *M. bovirhinis* and *M. alkalescens* are the major *Mycoplasma* species
causing mycoplasmal mastitis around the world. In Latin America the first report of a mastitis outbreak associated to *M. canadense* and *M. californicum* was described in Mexico; however, there is lack of information about its presence in other Latin American countries. In our case, strains B and C had been isolated from bulk tank milk from two different herds from Buenos Aires province (data not shown).

Volokhov *et al.* demonstrated that the ITS is a suitable and valuable marker for species identification among *Mollicutes*, since it showed a high percentage of interspecies diversity, low intraspecies variability and conserved flanking regions. Due to the labor-intensive and time-consuming isolation and biochemical identification of *Mycoplasma* species, their identification by amplification of the 16S-23S rRNA ITS region and further sequencing represents an easier and faster approach.

**Ethical responsibilities**

**Protection of human and animal subjects.** The authors declare that no experiments were performed on humans or animals for this study.

**Confidentiality of data.** The authors declare that no patient data appear in this article.

**Right to privacy and informed consent.** The authors declare that no patient data appear in this article.

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**Conflicts of interest**

The authors declare that they have no conflicts of interest.

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